

Nutritional Evaluation of Soybeans Varying in Trypsin Inhibitor Content¹

Y. HAN and C. M. PARSONS²

Department of Animal Sciences, University of Illinois, Urbana, Illinois 61801

T. HYMOWITZ

Department of Agronomy, University of Illinois, Urbana, Illinois 61801

(Received for publication August 23, 1990)

ABSTRACT A series of experiments was conducted to assess, by various methods, the nutritional value of a raw, Kunitz inhibitor-free, low trypsin-inhibitor soybean variant (LTS) in comparison with raw conventional soybeans (RCS) and heated dehulled soybean meal (HDS). The gross energy, protein, and amino acid concentrations of LTS were similar to those of RCS. The protein quality of the soybeans was compared in two trials in which young chicks were fed 9% or 16% CP diets containing one of the soybeans as the sole source of dietary protein. The protein quality of LTS was superior to that of RCS but inferior to that of HDS. True digestibilities of amino acids and TME_n were determined with a precision-fed cockerel assay using conventional and cecectomized birds. No significant effect of bird type on digestibility was found. The mean digestibility of 16 amino acids in RCS, LTS, and HDS was found to be approximately 68, 83, and 92%, respectively. The TME_n of LTS was greater than that of RCS. Chick growth assays indicated that amino acid bioavailability values of lysine and TSAA for LTS were greater than those for RCS but slightly lower than those for HDS.

In an additional chick assay with a corn and HDS diet (22% CP), the dietary HDS protein was replaced with LTS or RCS protein at 25, 50, 75, or 100%. Performance of chicks fed LTS was better than that of chicks fed RCS at all replacement levels, but the difference was not significant at the 25% replacement level. Feed efficiencies of chicks fed 25 or 50% of the dietary soybean protein as LTS were not significantly different from that of chicks fed the corn and HDS control diet. The results of the present study indicated that the nutritive value of LTS is substantially greater than that of RCS but somewhat lower than that of HDS. (*Key words:* soybeans, trypsin inhibitor, protein quality, amino acid, chicken)

1991 Poultry Science 70:896-906

INTRODUCTION

The detrimental effects on animal performance of trypsin inhibitors in raw soybeans have been extensively studied. Raw soybeans are nutritionally inferior to properly heated soybeans for rats (Osborne and Mendel, 1917; Lyman and Lepkovsky, 1957), chicks (Chernick *et al.*, 1948; Yen *et al.*, 1973) and pigs (Yen *et al.*, 1974; Cook *et al.*, 1988). The trypsin inhibitors in raw soybeans depress proteolytic activity in the small intestine, resulting in decreased release of free amino acids (AA). In addition, trypsin inhibitors cause pancreatic hypertrophy in rats and chicks due to stimulation of pancreatic secretions. The excess digestive enzymes subsequently se-

creted by the pancreas are largely undigested, resulting in increased AA excretion and deficiencies (Liener, 1977). Therefore, raw soybeans must be properly cooked in order to be maximally utilized by animals.

The Kunitz trypsin inhibitor is one of the major growth-inhibiting antinutritional factors present in raw soybeans (Rackis, 1965). A soybean variant low in Kunitz trypsin inhibitor activity was identified (Singh *et al.*, 1969; Clark and Hymowitz, 1972), and early studies showed that this variant was nutritionally superior to conventional commercial cultivars (Yen *et al.*, 1973). Recently, a new soybean variant has been developed at the University of Illinois that is isogenic to the commercially grown Williams 82 cultivar except that it lacks the Kunitz trypsin-inhibitor allele (Bernard and Hymowitz, 1986). Cook *et al.* (1988) reported that this variant was nutritionally superior to conventional raw soybeans for growing and finishing pigs. This new soybean variant might also be nutritionally and economically impor-

¹Supported in part by the Illinois Agricultural Experiment Station Project 20-0374 and by the Commodity Credit Corporation, Washington, DC.

²To whom reprint requests should be sent: 322 Mumford Hall, 1301 West Gregory Drive, Urbana, IL 61801.

tant to
of the
various
quality
trypsin
with]
Williar
vent-ex
Method
quality
oavaila
with c
to dete

Soybea

The
comme
were o
ment c
Illinois
Soybea
Experi
digesti
batch
RCS at
(L81-4
inhibiti
release
name]
similar
analysi
[Assoc
(AOAC
measur
strand
determ
Amino
(Spack
least tv
and Cy
mic ac
(1963)
remove
water.

Chick

One
cross o
Plymot
assays.
soybea

tant to the poultry industry. Thus, the objective of the present study was to evaluate, by various methods, and compare the protein quality of the raw Kunitz inhibitor-free, low-trypsin-inhibitor Williams soybeans (LTS) with protein quality of raw conventional Williams-82 soybeans (RCS) and heated solvent-extracted dehulled soybean meal (HDS). Methods of evaluation included chick protein quality assays, chick growth assays for bioavailable AA, and an excreta collection assay with conventional and cecectomized cockerels to determine true AA digestibility and TME_n .

MATERIALS AND METHODS

Soybean Sources

The HDS was purchased from a local commercial processing plant. The RCS and LTS were obtained in two batches from the Department of Agronomy, University of Illinois and Illinois Foundation Seeds, Inc., Champaign, IL. Soybeans from the first batch were used in chick Experiments 1, 2, 4, and 5 and in the true digestibility assay. Soybeans from the second batch were used in chick Experiment 3. The RCS and the LTS were an isolate of Williams 82 (L81-4590) that was lacking Kunitz trypsin-inhibitor activity. In 1990, L81-4590 was released for commercial production under the name Kunitz. All soybeans were ground to a similar particle size and subjected to proximate analysis for DM, N, ash, and gross energy [Association of Official Analytical Chemists (AOAC, 1980)]. Trypsin-inhibitor activity was measured according to the procedure of Hamerstrand *et al.* (1981), and urease activity was determined by the AOAC (1980) method. Amino acid concentrations were analyzed (Spackman *et al.*, 1958) on each sample with at least two replicates per sample. Analyses of Met and Cys were performed separately after performic acid oxidation by the method of Moore (1963), except the excess performic acid was removed by lyophilization after dilution with water.

Chick Assays

One-week-old male chicks resulting from the cross of New Hampshire males and Columbian Plymouth Rock females were used in six chick assays. The chicks were fed a 24% CP corn and soybean meal pretest diet during the first 7 days

posthatching. Following an overnight fast, chicks were weighed, wing-banded, and allotted to dietary treatment groups so that average body weights of chicks in each group were similar (Sasse and Baker, 1974). Triplicate groups of five or seven chicks were assigned to each dietary treatment, and the duration of each assay varied from 7 to 14 days. All of the chicks were housed in thermostatically controlled starter batteries with raised wire floors in an environmentally regulated room. Feed and water were supplied for *ad libitum* access, and uniform light was provided 24 h daily.

The first two chick experiments were conducted to determine and compare protein quality of soybean sources. In Experiment 1, chicks were fed a N-free basal diet (Willis and Baker, 1980) or the basal diet supplemented with one of the soybean sources to provide 9% dietary CP. The soybean sources were supplemented to the basal diet at the expense of cornstarch and dextrose. Protein efficiency ratio (PER) was calculated (in grams) as gain divided by CP intake. Net protein ratio (NPR) was calculated as [weight gain - weight gain of chicks fed N-free diet] divided by CP intake. Experiment 2 evaluated protein quality of the soybean sources

TABLE 1. Composition of diets used in Experiment 2 for assessing protein quality of heated dehulled soybean meal (HDS), raw conventional soybeans (RCS), and raw low-trypsin-inhibitor soybeans (LTS)¹

Ingredient	HDS	RCS	LTS
	%		
Soybean source	33.76	44.56	42.33
Cornstarch	27.74	21.94	24.08
Dextrose	26.50	26.50	26.50
Corn oil	5.0
Mineral premix ²	5.3	5.3	5.3
NaHCO ₃	1.0	1.0	1.0
Choline-Cl	.2	.2	.2
Vitamin premix ³	.2	.2	.2
DL-methionine	.3	.3	.3

¹Each diet contained 16% CP.

²Mineral mix provided per kilogram of diet: CaCO₃, 30.0 g; Ca₃(PO₄)₂, 28.0 g; K₂HPO₄, 9.0 g; NaCl, 8.8 g; MgSO₄·7H₂O, 3.5 g; MnSO₄·H₂O, .65 g; ferric citrate, .5 g; ZnCO₃, .1 g; CuSO₄·5H₂O, 20.0 mg; H₃BO₃, 9.0 mg; NaMoO₄·2H₂O, 9.0 mg; KI, 40.6 mg; CoSO₄·7H₂O, 1.0 mg; Na₂SeO₃, .215 mg.

³Supplied the following amounts per kilogram of diet: vitamin A, 4,400 IU; vitamin D₃, 1,000 ICU; vitamin E, 11 IU; vitamin B₁₂, .011 mg; riboflavin, 4.4 mg; D-pantothenic acid, 10 mg; niacin, 22 mg; menadione sodium bisulfate complex, 2.33 mg.

TABLE 2. Composition of diets used in Experiment 3¹

Ingredient	100 HDS ²	75 HDS:25 RCS	75 HDS:25 LTS	50 HDS:50 RCS	50 HDS:50 LTS	25 HDS:75 RCS	25 HDS:75 LTS	100 RCS	100 LTS
	(%)								
HDS	34.58	25.94	25.94	17.29	17.29	8.64	8.64
RCS	...	11.36	...	22.73	...	34.10	...	45.46	...
LTS	12.05	...	24.10	...	36.15	...	48.20
Ground corn (8.8% CP)	43.70	43.70	43.70	43.70	43.70	43.70	43.70	43.70	43.70
Fish meal (61% CP)	2.00	2.00	2.00	2.00	2.00	2.00	2.00	2.00	2.00
Alfalfa meal (17.5%)	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
Soybean oil	8.63	6.93	6.72	5.23	4.81	3.54	2.91	1.84	1.00
Cornstarch	2.08	1.67	1.56	1.26	1.04	.85	.52	.46	...
Dicalcium phosphate	2.20	2.20	2.20	2.20	2.20	2.20	2.20	2.20	2.20
Limestone	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
NaCl	.4	.4	.4	.4	.4	.4	.4	.4	.4
DL-methionine	.20	.20	.20	.20	.20	.20	.20	.20	.20
Choline-Cl	.10	.10	.10	.10	.10	.10	.10	.10	.10
Trace-mineral premix ³	.05	.05	.05	.05	.05	.05	.05	.05	.05
Vitamin premix ⁴	.10	.10	.10	.10	.10	.10	.10	.10	.10
Flavomycin	.05	.05	.05	.05	.05	.05	.05	.05	.05
Sand	1.96	1.65	1.47	1.35	.98	1.04	.49	.72	...
Wood cellulose	1.96	1.65	1.47	1.35	.98	1.04	.49	.72	...

¹All diets calculated to contain 22% total CP, 16.8% soybean CP and 8.4% digestible soybean oil using analytical values for the soybeans; CP (%), 48.47 for heated dehulled soybean meal (HDS), 36.92 for raw conventional soybeans (RCS) and 34.85 for raw Kunitz-free, low trypsin-inhibitor soybean meal (LTS); ether extract (%) 2.36 for HDS, 19.71 for RCS and 20.68 for LTS. The 100 HDS diet calculated to contain 3,125 kcal ME_N/kg.

²Numbers indicate percentages of dietary protein supplied by HDS, RCS, or LTS.

³Provided the following amounts per kilogram of diet: manganese, 75 mg; iron, 75 mg; zinc, 75 mg; copper, 5 mg; iodine, .75 mg; selenium, .1 mg.

⁴Supplied the following amounts per kilogram of diet: vitamin A, 4,400 IU; vitamin D₃, 1,000 ICU; vitamin E, 11 IU; vitamin B₁₂, .011 mg; riboflavin, 4.4 mg; d-pantothenic acid, 10 mg; niacin, 22 mg; menadione sodium bisulfate complex, 2.33 mg.

TABLE 4. Determination of protein quality of heated dehulled soybean meal (HDS), raw conventional soybeans (RCS), and raw low-trypsin-inhibitor soybeans (LTS) in Experiment 1¹

Treatment ²	Weight gain	Gain:feed	PER ³	NPR ³
	(g)	(g:g)		
Basal (N-free) ⁴	-7.2 ^d	-.104 ^d
Basal plus HDS	55.3 ^a	.343 ^a	3.81 ^a	4.31 ^a
Basal plus RCS	12.5 ^c	.121 ^c	1.21 ^c	2.16 ^c
Basal plus LTS	27.5 ^b	.227 ^b	2.52 ^b	3.18 ^b
Pooled SEM	2.3	.016	.19	.15

^{a-d}Means within columns with no common superscripts differ significantly ($P < .05$).

¹Means of three groups of five male chicks from 8 to 17 days posthatching.

²Each soybean source supplied 9% CP.

³PER [protein efficiency ratio (in grams)] = weight gain divided by protein intake. NPR [net protein ratio (in grams)] = [weight gain of birds fed a soybean diet minus weight gain of birds fed the N-free diet] divided by protein intake.

⁴Same as that used in protein quality assay II by Willis and Baker (1980).

True Digestibility Assay. Conventional (CONV) and cecectomized (CEC) Single Comb White Leghorn cockerels, 40 wk of age, were used to determine true digestibility of AA and TME_n in the various soybean sources. The birds were kept in individual cages with raised wire floors in an environmentally regulated room and provided with 16 h of light daily. Feed and water were supplied for *ad libitum* access before the onset of the experiment. Cecectomy was performed at 20 wk of age according to the procedure of Parsons (1985). Previous work in the authors' laboratory indicated that little or no cecal regrowth occurred in CEC cockerels within one year following the surgery (Parsons, 1985; Parsons, unpublished data). The CONV cockerels received a sham operation within a similar time period. The digestibility assay was the same as that described by Sibbald (1979), with some minor modifications. Following a 24-h fast, five CONV and five CEC cockerels were given 30 g of a test feedstuff via crop intubation. Five additional cockerels of each type were fasted throughout the experimental period to measure endogenous excretion of DM, energy, N, and AA. A plastic tray was placed under each cage, and excreta were collected for 48 h after crop intubation. Excreta samples were lyophilized, weighed, and ground to pass through a 60-mesh screen. Gross energy, N, and AA concentrations of each sample were analyzed by the procedures described previously.

Statistical Calculations and Analyses

True digestibilities were calculated for AA according to the method of Sibbald (1979) and

for TME_n by the method of Parsons *et al.* (1982). Statistical significance of differences in dry matter, TME_n, and true AA digestibility values were assessed with the Student's *t* test (Steel and Torrie, 1980).

Data from chick growth assays were subjected to analysis of variance procedures for completely randomized designs (Steel and Torrie, 1980). The least significant difference test based on the pooled SE from the analysis of variance was used for comparing treatment means. Slope-ratio methodology (Finney, 1978) was used to estimate bioavailability of AA in Experiments 4 to 6. Multiple regression equations were calculated with chick weight gain (in grams) as the dependent variable and the intake of supplemental test crystalline AA and feed-

TABLE 5. Effects of heated dehulled soybean meal (HDS), raw conventional soybeans (RCS), and raw low-trypsin-inhibitor soybeans (LTS) on performance of chicks fed a 16% protein diet supplemented with methionine in Experiment 2¹

Treatment	Weight gain	Gain:feed	PER ²
	(g)	(g:g)	
HDS	98.0 ^a	.632 ^a	3.95 ^a
RCS	42.9 ^c	.331 ^c	2.07 ^c
LTS	72.3 ^b	.489 ^b	3.06 ^b
Pooled SEM	2.0	.016	.10

^{a-c}Means within columns with no common superscripts differ significantly ($P < .05$).

¹Means of three groups of seven male crossbred chicks from 8 to 15 days posthatching.

²PER [Protein efficiency ratio (in grams)] = weight gain divided by protein intake.

stuff AA variables (millimo independ weight d each fec the ratio of the cry be 100% puted by gains of gain att crystallu ingredie 1986). T effect of among

The evaluate true dig 3. The c of RCS, protein these dit sources 2, Foot inhibito that in In the ment 1) gained i those fe chicks 1 that of c weight the LTS from tl observe

In E: diet co gained than di RCS (I perform that of Repl protein feed rat Experir of the d RCS or were n

w conventional soybeans
1¹

3 ³	NPR ³
a	4.31 ^a
c	2.16 ^c
b	3.18 ^b
	.15

et protein ratio (in grams)]
divided by protein intake.

Parsons *et al.* (1982).
f differences in dry
A digestibility values
dent's *t* test (Steel and

th assays were sub-
ance procedures for
signs (Steel and Tor-
ficant difference test
from the analysis of
omparing treatment
ology (Finney, 1978)
availability of AA in
ple regression equa-
chick weight gain (in
riable and the intake
alline AA and feed-

dehulled soybean meal
beans (RCS), and raw
LTS) on performance of
diet supplemented
Experiment 2¹

Gain:feed	PER ²
(g:g)	
.632 ^a	3.95 ^a
.531 ^c	2.07 ^c
.489 ^b	3.06 ^b
.016	.10

with no common super-
5).

en male crossbred chicks

io (in grams)] = weight

stuff AA (in milligrams) as the independent variables. In the TSAA assay, intake of sulfur (millimoles) from Met and Cys was used as the independent variable to account for molecular weight differences. The bioavailability of AA in each feedstuff was then estimated by means of the ratio of the slope of its regression line to that of the crystalline test AA, which was assumed to be 100% bioavailable. Bioavailability was computed by an additional method wherein weight gains of chicks were partitioned to reflect only gain attributable to intake of supplemental crystalline AA or supplemental AA from the test ingredient (Parsons, 1986; Hirakawa and Baker, 1986). This method attempts to minimize the effect of differences in voluntary feed intake among treatments.

RESULTS

The compositions of the soybean sources evaluated in Experiments 1, 2, 4, and 5 and the true digestibility assay are presented in Table 3. The composition of LTS was similar to that of RCS, although LTS was slightly higher in protein and lower in fat than RCS. However, these differences were reversed for the soybean sources evaluated in Experiment 3 (see Table 2, Footnote 1). The concentration of trypsin inhibitor activity in LTS was much lower than that in RCS and higher than that in HDS.

In the first protein quality assay (Experiment 1), chicks fed the diet containing LTS gained more than twice as much weight as did those fed RCS (Table 4). Feed efficiency of chicks fed the LTS diet was also superior to that of chicks fed the RCS diet. However, both weight gain and feed efficiency obtained from the LTS diet were inferior to those obtained from the HDS diet. Similar results were observed for PER and NPR values.

In Experiment 2, chicks fed a 16% protein diet containing LTS and supplemental Met gained weight more rapidly and efficiently than did chicks fed the same diet containing RCS (Table 5). As observed in Experiment 1, performance of chicks fed LTS was inferior to that of chicks fed HDS.

Replacing HDS with LTS or RCS in a 22% protein diet decreased weight gain and gain:feed ratio as the substitution level increased in Experiment 3 (Table 6). However, when 25% of the dietary soybean protein was provided by RCS or LTS, weight gain and gain:feed ratio were not significantly different ($P>.05$) from

TABLE 6. Effects of raw conventional soybeans (RCS) and raw low-trypsin-inhibitor soybeans (LTS) on chick performance in the soybean replacement assay in Experiment 3¹

Amount of soybean protein replaced	Weight gain	Feed intake	Gain:feed
	(g)		(g:g)
Control ²	223.5	323.0	.692
25% by RCS	216.9	322.9	.672
25% by LTS	219.7	324.0	.678
50% by RCS	203.6	322.5	.631
50% by LTS	211.6	319.0	.672
75% by RCS	194.7	317.0	.614
75% by LTS	208.2	320.5	.650
100% by RCS	174.9	317.4	.551
100% by LTS	198.6	319.4	.622
Pooled SEM	4.1	5.0	.008

¹Means of three groups of seven male crossbred chicks from 8 to 19 days posthatching.

²Control diet was a 22% CP corn and soybean meal diet in which 100% of the dietary soybean protein was supplied by heated, dehulled soybean meal.

those of the control. In addition, feed efficiency of chicks provided 50% of the soybean protein as LTS was not significantly different ($P>.05$) from that of the control. At each replacement level, chick performance was better on LTS diets than on RCS diets, with the differences being significant ($P<.05$) at the 75 and 100% substitution rates. Regressing weight gain (in grams) on percentage replacement level of HDS protein by RCS or LTS protein yielded the regression equation: $Y = 225.6 - .4644 (RCS) - .2592 (LTS)$, $R^2 = .81$, indicating that each 1% replacement of HDS protein with RCS protein depressed weight gain by .46 g and each 1% replacement of HDS protein with LTS protein depressed weight gain by .26 g. The slope ratio of LTS:RCS was .558. When regressing gain:feed ratio on the same substitution levels, the equation was $Y = .697 - .001319 (RCS) - .0007076 (LTS)$, $R^2 = .90$, and the slope ratio of LTS:RCS was .536.

Weight gain and gain:feed ratio responded linearly to supplementation of the AA-deficient basal diets with crystalline AA or the soybean sources in the AA bioavailability assays (Tables 7 to 9). From multiple linear regression analysis and slope-ratio methodology applied to total weight gains, bioavailability of Lys was ($\bar{x} \pm SE$) $90.4 \pm 5.0\%$ for HDS, $72.0 \pm 6.1\%$ for RCS, and $86.4 \pm 6.0\%$ for LTS in Experiment 4 (Table 7). With parti-

TABLE 7. Determination of lysine bioavailability in heated dehulled soybean meal (HDS), raw conventional soybeans (RCS), and raw low-trypsin-inhibitor soybeans (LTS) by slope-ratio chick assay in Experiment 4¹

Treatment	Weight gain		Gain:feed	Bioavailability ²	
	Total	Part. ³		Total	Part. ³
	— (g per chick) —		(g:g)	———— (%) ————	
1. Basal (B) ⁴	36.0	-1.6	.360		
2. B plus .15% L-Lys	66.3	19.0	.517		
3. B plus .30% L-Lys	102.4	41.5	.608	100.0	100.0
4. B plus 3% HDS	46.8	9.0	.465		
5. B plus 6% HDS	64.1	19.6	.534		
6. B plus 9% HDS	83.1	35.0	.595	90.4 ± 5.0	101.8 ± 6.5
7. B plus 3% RCS	40.5	3.6	.414		
8. B plus 6.5% RCS	56.7	15.6	.516		
9. B plus 10% RCS	63.9	18.6	.523	72.0 ± 6.1	80.4 ± 7.7
10. B plus 3% LTS	48.5	7.2	.437		
11. B plus 6.5% LTS	56.1	14.0	.496		
12. B plus 10% LTS	72.4	24.6	.559	86.4 ± 6.0	92.6 ± 7.6
Pooled SEM	2.8	1.9	.014		

¹Values are means of triplicate groups of five male chicks from 8 to 16 days posthatching.

² $\bar{x} \pm SE$ calculated from slope-ratio analysis for multiple regression of total or partitioned gain (grams) on supplemental crystalline lysine or soybean lysine intake (milligrams); the regression equations were $Y = 36.72 + .1364(L-Lys) + .1233(HDS) + .1178(LTS) + .09825(RCS)$, $R^2 = .96$ and $Y = -.48 + .08511(L-Lys) + .08667(HDS) + .06840(RCS) + .07882(LTS)$, $R^2 = .94$, respectively.

³Part. = Partitioned gain or bioavailability, which reflects gain or bioavailability due only to supplemental crystalline amino acid or soybean amino acid intake. The reference standard curve from regressing total gain (Y; in grams) on total lysine intake (X; in milligrams) for Treatments 1 to 3 was $Y = 3.61 + .08641X$, $r^2 = .99$.

⁴Purified crystalline amino acid diet deficient only in lysine (Lys; .4% L-Lys).

TABLE 8. Determination of total sulfur amino acid bioavailability in heated dehulled soybean meal (HDS), raw conventional soybeans (RCS), and raw low-trypsin-inhibitor soybeans (LTS) by slope-ratio chick assay in Experiment 5¹

Treatment	Weight gain		Gain:feed	Bioavailability ²	
	Total	Part. ³		Total	Part. ³
	— (g per chick) —		(g:g)	———— (%) ————	
1. Basal (B) ⁴	27.5	-2.0	.277		
2. B plus .035% L-Cys + .035% L-Met	55.9	13.5	.424		
3. B plus .07% L-Cys + .07% L-Met	72.1	26.2	.511	100.0	100.0
4. B plus 4% HDS	43.1	4.9	.358		
5. B plus 8% HDS	66.3	22.6	.490		
6. B plus 12% HDS	79.9	31.7	.543	89.2 ± 6.4	92.2 ± 8.1
7. B plus 4% RCS	38.2	4.0	.344		
8. B plus 8% RCS	42.3	10.1	.400		
9. B plus 12% RCS	51.2	15.7	.448	62.3 ± 7.7	82.5 ± 9.6
10. B plus 4% LTS	40.0	5.6	.358		
11. B plus 8% LTS	52.1	13.3	.424		
12. B plus 12% LTS	61.9	19.2	.466	86.6 ± 8.0	90.0 ± 10.2
Pooled SEM	2.7	2.1	.016		

¹Values are means of triplicate groups of five male chicks from 8 to 16 days posthatching.

² $\bar{x} \pm SE$ calculated from slope-ratio analysis for multiple regression of total or partitioned gain (grams) on supplemental sulfur intake from crystalline TSAA or soybean TSAA (millimoles); the regression equations were $Y = 30.63 + 30.11(TSAA) + 26.86(HDS) + 18.76(RCS) + 26.05(LTS)$, $R^2 = .93$ and $Y = -1.31 + 19.27(TSAA) + 17.76(HDS) + 15.89(RCS) + 17.34(LTS)$, $R^2 = .90$, respectively.

³Part. = partitioned gain or bioavailability, which reflects gain or bioavailability due only to supplemental crystalline amino acid or soybean amino acid intake. The reference standard curve from regressing total gain (Y; in grams) on intake of sulfur from TSAA (X; in millimoles) for Treatments 1 to 3 was $Y = -9.35 + 28.88X$, $r^2 = .99$.

⁴Purified crystalline amino acid diet deficient only in TSAA (.125% L-Met, .125% L-Cys).

TABLE
soyl

Treatme

- 1. Bas
- 2. B f
- 3. B f
- 4. B f
- 5. B f
- 6. B f
- 7. B f
- 8. B f
- 9. B f
- 10. B f
- 11. B f
- 12. B f

¹Val

²Par

amino a
methion

³ $\bar{x} \pm$

supplem

Met) + .

.2148(L

⁴Pur

tioned

oavaila

corresp

weight

not sig

Bio

total w

and 86

respect

bioavai

tioned

total

signific

The

of the l

much l

HDS, v

diate i

bioavai

LTS (6

in HDS

than th

derived

among

manner

True

values

TABLE 9. Determination of methionine bioavailability in heated dehulled soybean meal (HDS), raw conventional soybeans (RCS), and raw low-trypsin-inhibitor soybeans (LTS) by slope-ratio chick assay in Experiment 6¹

Treatment	Weight gain		Gain:feed	Bioavailability ³	
	Total	Part. ²		Total	Part. ²
	— (g per chick) —		(g:g)	— (%) —	
1. Basal (B) ⁴	18.1	-1.4	.262		
2. B plus .05% L-Met	55.1	19.0	.489		
3. B plus .10% L-Met	111.7	51.1	.633	100.0	100.0
4. B plus 4% HDS	31.6	5.5	.364		
5. B plus 9% HDS	45.7	15.9	.475		
6. B plus 14% HDS	67.0	28.4	.563	80.3 ± 4.0	87.3 ± 4.9
7. B plus 5% RCS	23.5	2.6	.322		
8. B plus 10% RCS	29.7	5.3	.362		
9. B plus 15% RCS	42.7	11.1	.424	55.4 ± 5.7	51.4 ± 7.3
10. B plus 5% LTS	26.6	3.4	.336		
11. B plus 10% LTS	33.3	6.5	.377		
12. B plus 15% LTS	48.4	17.7	.495	67.9 ± 5.7	72.1 ± 6.9
Pooled SEM	3.4	2.1	.015		

¹Values are means of triplicate groups of five male chicks from 8 to 16 days posthatching.

²Part. = Partitioned gain or bioavailability, which reflects gain or bioavailability due only to supplemental crystalline amino acid or soybean amino acid intake. The reference standard curve from regressing total gain (Y; in grams) on total methionine (Met) intake (X; in milligrams) for Treatments 1 to 3 was $Y = -7.11 + .2956X$, $r^2 = .995$.

³ $\bar{x} \pm SE$ calculated from slope-ratio analysis for multiple regression of total or partitioned gain (grams) on supplemental crystalline Met or soybean Met intake (milligrams); the regression equations were $Y = 19.64 + .5338(L-Met) + .4287(HDS) + .2955(RCS) + .3625(LTS)$, $R^2 = .98$ and $Y = -.43 + .2978(L-Met) + .2600(HDS) + .1531(RCS) + .2148(LTS)$, $R^2 = .97$, respectively.

⁴Purified crystalline amino acid diet deficient only in Met (.13% L-Met; .30% L-Cys).

tioned weight gains, the estimated Lys bioavailabilities were slightly higher than the corresponding values obtained from total weight gains; however, the differences were not significant ($P > .05$).

Bioavailabilities of TSAA calculated from total weight gains were 89.2 ± 6.4 , 62.3 ± 7.7 , and $86.6 \pm 8.0\%$ for HDS, RCS, and LTS, respectively, in Experiment 5 (Table 8). The bioavailability estimate computed from partitioned gain was similar to that computed from total gain for HDS and LTS, but was significantly higher for RCS ($82.5 \pm 9.6\%$).

The response of chicks to supplementation of the MET-deficient basal diet with RCS was much lower than that to supplementation with HDS, with the response to LTS being intermediate in Experiment 6 (Table 9). The Met bioavailability based on total weight gain in LTS ($67.9 \pm 5.7\%$) was lower ($P < .05$) than that in HDS ($80.3 \pm 4.0\%$) but was greater ($P < .05$) than that in RCS ($55.4 \pm 5.7\%$). The estimates derived from partitioned weight gains differed among the three soybean sources in a similar manner.

True DM and AA digestibility and TME_n values are presented in Table 10. There were

no significant ($P > .05$) differences between CONV and CEC cockerels for true digestibility of DM and AA within soybean sources except for Tyr in HDS. True digestibility values for AA in HDS were significantly ($P < .05$) greater than all of those in RCS and most of those in LTS. True AA digestibility values for LTS were greater ($P < .05$) than those for RCS. True digestibility of DM in LTS was greater than that in HDS. As expected, TME_n values of RCS and LTS were greater ($P < .05$) than that of HDS, and TME_n of LTS was greater ($P < .05$) than TME_n of RCS. The TME_n of RCS determined with CEC birds was greater than the TME_n determined with CONV birds, whereas the reverse was observed for LTS.

DISCUSSION

Proper heating of soybeans improves their nutritive value to animals by destroying trypsin inhibitors (Kunitz, 1947; Rackis *et al.*, 1962) and hemagglutinin (Liener, 1951, 1953). The chick protein quality assays (Experiments 1 and 2) clearly illustrated the benefits of

TABLE 10. True metabolizable energy and true digestible amino acid values for heated dehulled soybean meal (HDS), raw conventional soybeans (RCS), and raw low-trypsin-inhibitor soybeans (LTS)¹

	HDS			RCS			LTS		
	CONV ²	CEC ²	SED ²	CONV	CEC	SED	CONV	CEC	SED
True digestibility of DM, %	45.7	41.6	3.8	48.4	48.7	3.3	54.9	53.9	4.1
TME _n , kcal/g DM	2.742	2.881	.098	3.311 ^a	3.623 ^b	.065	3.944 ^a	3.751 ^b	.082
True digestibility of amino acids, %									
Aspartic acid	91.8	90.2	1.2	68.7	69.5	2.3	85.2	80.0	4.4
Threonine	90.3	88.8	1.6	61.8	63.3	2.5	82.7	76.7	4.3
Serine	92.2	92.8	1.0	66.4	67.9	2.4	85.5	79.8	4.1
Glutamic acid	94.4	95.3	1.0	76.6	81.0	2.7	88.9	86.8	4.2
Proline	94.6	94.1	1.1	72.7	76.2	2.3	86.8	82.2	3.9
Alanine	88.1	89.9	2.1	60.8	64.8	3.1	80.6	78.3	5.6
Cystine	85.5	86.2	1.4	57.0	51.9	5.1	79.1	69.3	5.0
Valine	93.1	93.5	1.8	61.7	68.3	5.8	83.1	79.6	6.4
Methionine	93.1	90.4	2.8	62.5	56.2	5.8	81.2	81.1	6.7
Isoleucine	94.2	95.0	1.2	66.4	72.2	4.7	84.9	83.7	5.5
Leucine	93.2	93.7	1.1	67.0	70.0	2.7	85.2	82.4	4.6
Tyrosine	93.2 ^a	96.7 ^b	1.3	64.1	63.1	2.1	84.5	77.3	4.6
Phenylalanine	94.7	96.2	1.1	67.4	70.8	3.2	86.6	82.4	5.5
Histidine	93.6	94.0	.7	73.7	70.6	5.0	86.5	87.0	5.7
Lysine	92.7	93.9	1.0	76.0	78.6	2.9	86.8	85.6	3.6
Arginine	92.0	91.9	2.2	75.5	79.9	2.7	88.7	86.9	3.9
\bar{x}	92.3	92.7		67.4	69.0		84.8	81.2	

^{a,b}Within soybean source, means in the same row with no common superscripts are significantly different ($P < .05$).

¹Values are means of five birds.

²CONV = Conventional birds; CEC = cecectomized birds; SED = standard error of the difference.

cooking
trypsin
factors
efficient
RCS w
PER fr
that fr
that fr
with N
and LT
Exper
suppl
correct
AA fr
precus
and, t
from t
1977).
The
indicat
to RC
agree
Yen et
soybea
with r
(1988)
cultiva
LTS to
that K
two m
inhibit
Hymo
(1973)
depres
accou
heatin
soybe
pancre
ments
its pr
1990;
Th
true c
AA a
for th
comp.
and 1
gener
than 1
proba
endog
enzym
biliary
activa
The 1

cooking soybeans and the deleterious effects of trypsin inhibitors and other antinutritional factors in raw soybeans. For example, the efficiency of protein utilization by chicks fed RCS was only 32% of those fed HDS. The PER from LTS was substantially superior to that from RCS but was still 34% lower than that from HDS. When diets were supplemented with Met, PER was improved for both RCS and LTS relative to HDS (Experiment 2 versus Experiment 1). The positive response to Met supplementation was probably due both to the correction of the deficiency of the first-limiting AA for growth and to the provision of precursor Cys for pancreatic enzyme synthesis, and, thus, reduction of the depletion of Cys from body tissues (Yen *et al.*, 1973; Liener, 1977).

The results of the protein quality assays indicated that LTS was nutritionally superior to RCS but inferior to HDS. These results agree with results of the early chick study of Yen *et al.* (1973) using low trypsin inhibitor soybeans of varying genetic background and with results of the pig study of Cook *et al.* (1988) using the same commercial Williams cultivars evaluated herein. The inferiority of LTS to HDS may be partially due to the fact that Kunitz trypsin inhibitor is only one of the two major classes of trypsin and chymotrypsin inhibitors present in soybeans (Stahlhut and Hymowitz, 1983). In addition, Kakade *et al.* (1973) reported that only 40% of the growth-depressing effect of raw soybeans could be accounted for by trypsin inhibitors and that heating is necessary for denaturation of the soybean proteins and subsequent digestion by pancreatic enzymes. Further, recent experiments indicate that heating of LTS increases its protein quality markedly (Friedman *et al.*, 1990; Parsons *et al.*, unpublished data).

The chick AA bioavailability assays and the true digestibility assay indicated that reduced AA availability is at least partially responsible for the lower protein quality of LTS and RCS compared with that of HDS. Bioavailability and true digestibility of AA in LTS were generally higher than those in RCS but lower than those in HDS. The lower AA digestibility probably resulted both from increased loss of endogenous protein due to increased pancreatic enzyme secretions and from decreased digestibility of dietary AA due to reduced proteolytic activity (Bielorai *et al.*, 1973; Liener, 1977). The higher TME_n of LTS compared with RCS

was probably largely due to the higher availability of the AA in LTS.

True digestibility values for Lys determined by excreta collection assay with adult birds agreed very well with the bioavailability values determined by growth assay with chicks. However, digestible Met values were somewhat higher than bioavailable Met values for HDS and LTS, whereas the values were in good agreement for RCS. Calculation method (total weight gain versus partitioned weight gain) had little effect on chick growth assay bioavailability values in the present study.

It has been reported that there is a significant effect of the hindgut microflora on AA digestibility in some feedstuffs for poultry (Parsons, 1986). However, cecectomy had no significant effect on AA digestibility values for any of the soybean sources tested in the present study. In contrast, digestibility values for AA in RCS in ileal-cannulated pigs were reported to be markedly lower than digestibility values for CONV pigs (Vandergrift *et al.*, 1983). These observations suggest that digestion and metabolism of raw soybean protein by the hindgut microflora is much less in poultry than in swine.

The results of the soybean protein replacement assay showed that the growth depression caused by replacing HDS with LTS was only approximately one-half of that caused by replacing HDS with RCS. These results clearly show the nutritional superiority of HDS compared to RCS when used in a practical-type diet. Moreover, weight gain and feed efficiency of chicks fed the diets containing 25 or 50%, respectively, of the soybean protein from LTS were not significantly different from those of chicks fed a diet containing 100% of the soybean protein from HDS. It is also possible that even higher levels of LTS could be used in diets of older birds without adversely affecting performance, as the adverse response to dietary raw soybeans is, to some degree, age-dependent (Crenshaw and Danielson, 1985).

REFERENCES

- Adams, K. L., and A. H. Jensen, 1984. Comparative utilization of inseed fats and the respective extracted fats by the young pig. *J. Anim. Sci.* 59:1557-1566.
- Association of Official Analytical Chemists, 1980. *Official Methods of Analysis*. 13th ed. Association of Official Analytical Chemists, Washington, DC.
- Baker, D. H., K. R. Robbins, and J. J. Buck, 1979.

¹Values are means of five birds.

²CONV = Conventional birds; CEC = cecectomized birds; SED = standard error of the difference.

- Modification of the level of histidine and sodium bicarbonate in the Illinois crystalline amino acid diet. *Poultry Sci.* 58:749-750.
- Bernard, R. L., and T. Hymowitz, 1986. Registration of L81-4871, and L83-4387 soybean germplasm lines lacking the Kunitz trypsin inhibitor. *Crop Sci.* 26: 650-651.
- Bielorai, R., M. Tamir, E. Alumot, A. Bar, and S. Hurwitz, 1973. Digestion and absorption of protein along the intestinal tract of chicks fed raw and heated soybean meal. *J. Nutr.* 103:1291-1298.
- Chemick, S. S., S. Lepkovsky, and I. L. Chaikoff, 1948. A dietary factor regulating the enzyme content of the pancreas: changes induced in size and proteolytic activity of the chick pancreas by the ingestion of raw soybean meal. *Am. J. Physiol.* 155:34-41.
- Clark, R. W., and T. Hymowitz, 1972. Activity variation between and within two soybean trypsin inhibitor electrophoretic forms. *Biochem. Genet.* 6:169-182.
- Cook, D. A., A. H. Jensen, J. R. Farley, and T. Hymowitz, 1988. Utilization by growing and finishing pigs of raw soybeans of low Kunitz trypsin inhibitor content. *J. Anim. Sci.* 66:1686-1691.
- Crenshaw, M. A., and D. M. Danielson, 1985. Raw soybeans for growing-finishing pigs. *J. Anim. Sci.* 60:725-730.
- Finney, D. J., 1978. *Statistical Method of Biological Assay*. 3rd ed. Charles Griffin & Co. Ltd., High Wycombe, Buckinghamshire, England.
- Friedman, M., D. L. Brandon, A. H. Bates, and T. Hymowitz, 1990. Comparison of a commercial/soybean cultivar and an isolate lacking the Kunitz trypsin inhibitor: Composition, nutritional value, and effects of heating. *J. Agric. Food. Chem.* (in press).
- Hamerstrand, G. E., L. T. Black, and J. D. Glover, 1981. Trypsin inhibitors in soy products: Modification of the standard analytical procedure. *Cereal Chem.* 58: 42-45.
- Hirakawa, D. A., and D. H. Baker, 1986. Assessment of lysine bioavailability of an intact protein mixture: Comparison of chick growth and precision-fed rooster assays. *Nutr. Res.* 6:815-826.
- Kakade, M. L., D. E. Hoffa, and I. E. Liener, 1973. Contribution of trypsin inhibitors to the deleterious effects of unheated soybeans fed to rats. *J. Nutr.* 103: 1772-1778.
- Kunitz, M. 1947. Crystalline trypsin inhibitor. II. General properties. *J. Gen. Physiol.* 30:291-310.
- Liener, I. E., 1951. The intraperitoneal toxicity of concentrates of the soybean trypsin inhibitor. *J. Biol. Chem.* 193:183-191.
- Liener, I. E., 1953. Soyin, a toxic protein from the soybean. I. Inhibition of rat growth. *J. Nutr.* 49: 527-539.
- Liener, I. E., 1977. Protease inhibitors and hemagglutinins of legumes. Pages 284-303 in: *Evaluation of Proteins for Humans*. C. E. Bodwell, ed. The AVI Publishing Company, Inc., Westport, CT.
- Lyman, R. L., and S. Lepkovsky, 1957. The effect of raw soybean meal and trypsin inhibitor diets on pancreatic enzyme secretion in the rat. *J. Nutr.* 62:269-284.
- Moore, S., 1963. On the determination of cystine as cysteic acid. *J. Biol. Chem.* 238:235-237.
- Osborne, T. B., and L. B. Mendel, 1917. The use of soybean as food. *J. Biol. Chem.* 32:369-387.
- Parsons, C. M., 1985. Influence of caecectomy on digestibility of amino acids by roosters fed distiller's dried grains with solubles. *J. Agric. Sci. Camb.* 104: 496-472.
- Parsons, C. M., 1986. Determination of digestible and available amino acids in meal using conventional and caecotomized cockerels or chick growth assays. *Brit. J. Nutr.* 56:227-240.
- Parsons, C. M., L. M. Potter, and B. A. Bliss, 1982. True metabolizable energy corrected to nitrogen equilibrium. *Poultry Sci.* 61:2241-2246.
- Rackis, J. J., 1965. Physiological properties of soybean trypsin inhibitors and their relationship to pancreatic hypertrophy and growth inhibition of rats. *Fed. Proc.* 24:1488-1493.
- Rackis, J. J., H. A. Sasame, R. K. Mann, R. L. Anderson, and A. K. Smith, 1962. Soybean trypsin inhibitors: Isolation, purification and physical properties. *Arch. Biochem. Biophys.* 78:471-478.
- Sasse, C. E., and D. H. Baker, 1974. Factors affecting sulfate-sulfur utilization by the young chick. *Poultry Sci.* 53:652-662.
- Sibbald, I. R., 1979. A bioassay for available amino acids and true metabolizable energy in feedstuffs. *Poultry Sci.* 58:668-673.
- Singh, L., C. M. Wilson, and H. H. Hadley, 1969. Genetic differences in soybean trypsin inhibitors separated by disc electrophoresis. *Crop Sci.* 9:489-491.
- Spackman, D. H., W. H. Stein, and S. Moore, 1958. Automatic recording apparatus for use in the chromatography of amino acids. *Anal. Chem.* 30: 1190-1206.
- Stahlhut, R. W., and T. Hymowitz, 1983. Variation in the low molecular weight proteinase inhibitors of soybeans. *Crop Sci.* 23:766-769.
- Steel, R.G.D., and J. H. Torrie, 1980. *Principles and Procedures of Statistics. A Biometrical Approach*. 2nd ed. McGraw-Hill Book Co., Inc., New York, NY.
- Vandergrift, W. L., D. A. Knabe, T. D. Tanksley, Jr., 1983. Digestibility of nutrients in raw and heated soyflakes for pigs. *J. Anim. Sci.* 57:1215-1224.
- Willis, G. M., and D. H. Baker, 1980. Evaluation of turfgrass clippings as a dietary ingredient for the growing chick. *Poultry Sci.* 59:404-411.
- Yen, J. T., T. Hymowitz, and A. H. Jensen, 1974. Effects of soybeans of different trypsin-inhibitor activities on performance of growing swine. *J. Anim. Sci.* 38: 304-309.
- Yen, J. T., A. H. Jensen, T. Hymowitz, and D. H. Baker, 1973. Utilization of different varieties of raw soybeans by male and female chicks. *Poultry Sci.* 52: 1875-1882.

ABSTR.
MB; lo
increas
deficien
by sele
Laying
Dui
energy
after 16
product
birds w
1,140 g
perform
(Key w

The nutr
ment pullet
literature v
mendations
controlled
during the
regimens (I
al., 1981;
regimens (I
et al., 1983
single-stage
Summers,
all been us
The effe
rearing per
ance is no
that the en
on laying p
rison, 1977
showed a s
and feed cc
(Wolf et a
Few reg
high-densit

¹Published
Series, Minne
²To whom